To detect gene-environment interactions, a logistic regression model is typically fitted to a set of case-control data, and the focus is on testing of the cross-product terms (gene × environment) in the model. A significant result is indicative of a gene-environment interaction under a multiplicative model for disease odds. Based on the sufficient-cause model for rates, in this paper we put forward a general approach to testing for sufficient-cause gene-environment interactions in case-control studies. The proposed tests can be tailored to detect a particular type of sufficient-cause gene-environment interaction with greater sensitivity. These tests include testing for autosomal dominant, autosomal recessive, and gene-dosage interactions. The tests can also detect trend interactions (e.g., a larger gene-environment interaction with a higher level of environmental exposure) and threshold interactions (e.g., gene-environment interaction occurs only when environmental exposure reaches a certain threshold level). Two assumptions are necessary for the validity of the tests: 1) the rare-disease assumption and 2) the no-redundancy assumption. Another 2 assumptions are optional but, if imposed correctly, can boost the statistical powers of the tests: 3) the gene-environment independence assumption and 4) the Hardy-Weinberg equilibrium assumption. SAS code (SAS Institute, Inc., Cary, North Carolina) for implementing the methods is provided.

The occurrence of most human diseases is the result of interplay between genetic and environmental factors (1, 2). To detect gene-environment interactions, epidemiologists often adopt a case-control study design, recruiting diseased subjects (cases) and nondiseased subjects (controls) and comparing their genotypes and environmental exposures. A logistic regression model is typically fitted to the data, and the focus is on hypothesis testing of the cross-product terms (gene × environment) in the model (3). A significant result is indicative of a gene-environment interaction under a multiplicative model for disease odds. Oftentimes, it is reasonable to assume that the genes under examination are in Hardy-Weinberg equilibrium (HWE) and are also independent of any environmental exposure among the nondiseased subjects in the study population (4–9). A modified logistic regression methodology developed by Lee et al. (9) can exploit these 2 assumptions and achieve higher statistical powers for the multiplicative interaction tests.

Based on the sufficient-cause model for risks, VanderWeele and Robins (10, 11) constructed statistical tests for causal mechanistic interactions between binary variables. Basically, these are additive interaction tests, examining whether the observed disease risks deviate too much from additivity. VanderWeele (12) then went on to expand the methodologies for including categorical or ordinal variables with 3 or more levels. This later development is of particular relevance to the
current study of detecting gene-environment interactions, because genes are often coded as ternary variables with levels indicating 0, 1, or 2 variant alleles; the environmental exposures under study can also have multiple levels.

Based on the sufficient-cause model for rates, Lee (13, 14) constructed statistical tests for causal mechanistic interactions between binary variables and determined that a rate-model–based test has a less stringent threshold for detecting causal mechanistic interactions than a corresponding risk-model–based test. In this paper, I build upon previous work (13, 14) to propose a general hypothesis-testing framework for sufficient-cause gene-environment interactions. The proposed tests can be tailored to detect a particular type of sufficient-cause gene-environment interaction with greater sensitivity. These include testing for autosomal dominant, autosomal recessive, or gene-dosage interactions. The tests can also detect trend interactions (e.g., a larger gene-environment interaction with a higher level of environmental exposure) and threshold interactions (e.g., gene-environment interaction that occurs only when the environmental exposure reaches a certain threshold level). Two assumptions are necessary for the validity of these tests: 1) the rare-disease assumption and 2) the no-redundancy assumption (13–15). Another 2 assumptions are optional but, if imposed correctly, can boost the statistical powers of the tests: 3) the gene-environment independence (GEI) assumption and 4) the HWE assumption.

METHODS

Sufficient-cause model for rates

Let \( G \) (\( G = 0, 1, \) or \( 2 \)) represent the number of variant alleles a subject carries and \( E \) (\( E = 0 \) or \( 1 \)) represent a binary environmental exposure. Let \( \text{Rate}_{g,e} \) (Odds \( g,e \)) denote the disease rate (odds) for subjects with \( G = g \) and \( E = e \) in the study population. The rare-disease assumption is invoked, so disease odds (for 1 unit of follow-up time) and disease rates are equivalent (3).

The above variables \((G \) and \( E \)) together define a total of 12 classes of sufficient causes (i.e., \( (3 + 1) \times (2 + 1) = 12 \)), including 1 “all-unknown” class \((U_1)\), 3 gene-only classes \((U_2, U_3, U_4)\), 2 environment-only classes \((U_5, U_6)\), and 6 gene-environment interaction classes \((U_7 \sim U_{12})\) (Figure 1). Note that here we do not impose the assumption of monotonicity (16–20) on the genetic effect, the environmental effect, or the gene-environment interaction effect, so this represents the most general sufficient-cause model for a ternary \( G \) and a binary \( E \).

The sufficient-cause model is partly deterministic and partly stochastic. The presence of risk factor(s) alone is not sufficient for the disease. Only when all of the unknown components (complement causes) also appear can the sufficient cause become complete and the disease occur. Let \( \text{Rate}_{U_{12}} \sim \text{Rate}_{U_{12j}} \) denote the “completion rates” of the aforementioned 12 classes of sufficient causes, respectively. The completion rate for a particular class is the instantaneous arrival rate of the unknown complement causes in that class (20). Here the no-redundancy assumption is applied. This assumption posits that within a sufficiently short time interval, there can only be, at most, 1 arrival event of the unknown complement causes for each and every subject in the population (13–15).

Under the no-redundancy assumption, the disease rate (and also the disease odds under the rare-disease assumption) for subjects with \( G = g \) and \( E = e \) is a simple arithmetic sum of the completion rates of the 4 “completable classes” (13, 14), that is,

\[
\begin{align*}
\text{Odds}_{2,1} &= \text{Rate}_{2,1} = \text{Rate}_{U_1} + \text{Rate}_{U_2} + \text{Rate}_{U_3} + \text{Rate}_{U_4}, \\
\text{Odds}_{1,1} &= \text{Rate}_{1,1} = \text{Rate}_{U_1} + \text{Rate}_{U_2} + \text{Rate}_{U_3} + \text{Rate}_{U_4}, \\
\text{Odds}_{0,1} &= \text{Rate}_{0,1} = \text{Rate}_{U_1} + \text{Rate}_{U_4} + \text{Rate}_{U_5} + \text{Rate}_{U_6}, \\
\text{Odds}_{2,0} &= \text{Rate}_{2,0} = \text{Rate}_{U_1} + \text{Rate}_{U_2} + \text{Rate}_{U_3} + \text{Rate}_{U_{10}}, \\
\text{Odds}_{1,0} &= \text{Rate}_{1,0} = \text{Rate}_{U_1} + \text{Rate}_{U_3} + \text{Rate}_{U_5} + \text{Rate}_{U_{11}},
\end{align*}
\]

Figure 1. The 12 total classes of sufficient component causes for a ternary gene \((G = 0, 1, \text{ or } 2)\) and a binary environmental exposure \((E = 0 \text{ or } 1)\): the all-unknown class \((U_1)\), the gene-only classes \((U_2, U_3, U_4)\), the environment-only classes \((U_5, U_6)\), and the gene-environment interaction classes \((U_7 \sim U_{12})\).
and
\[
\text{Odds}_{0,0} = \text{Rate}_{0,0} = \text{Rate}_{U_1} + \text{Rate}_{U_4} + \text{Rate}_{U_8} + \text{Rate}_{U_{12}},
\]
respectively. In the case of subjects with \( G = 2 \) and \( E = 1 \), for example, they can develop the disease because of the completion of the \( U_1, U_2, U_5, \) or \( U_7 \) class; these 4 classes are their compatible classes.

### Testing for sufficient-cause gene-environment interactions

Now, consider the following interaction contrasts (ICs) which quantify departure from additivity:

\[
\text{IC}_1 = \text{Odds}_{s,1} - \text{Odds}_{s,0} - \text{Odds}_{s,0} + \text{Odds}_{s,0} \\
= \text{Rate}_{U_8} - \text{Rate}_{U_9} - \text{Rate}_{U_{11}} + \text{Rate}_{U_{12}},
\]

\[
\text{IC}_2 = \text{Odds}_{s,1} - \text{Odds}_{s,0} - \text{Odds}_{s,2} + \text{Odds}_{s,0} \\
= \text{Rate}_{U_4} - \text{Rate}_{U_9} - \text{Rate}_{U_{10}} + \text{Rate}_{U_{12}},
\]

and

\[
\text{IC}_3 = w_1 \times \text{IC}_1 + w_2 \times \text{IC}_2 \\
= [w_2 \times \text{Odds}_{s,1} + w_1 \times \text{Odds}_{s,1} - (w_2 + w_1) \times \text{Odds}_{s,0}] \\
- [w_2 \times \text{Odds}_{s,0} + w_1 \times \text{Odds}_{s,0} - (w_2 + w_1) \times \text{Odds}_{s,0}] \\
= [w_2 \times \text{Rate}_{U_8} + w_1 \times \text{Rate}_{U_8} - (w_2 + w_1) \times \text{Rate}_{U_8}] \\
- [w_2 \times \text{Rate}_{U_{10}} + w_1 \times \text{Rate}_{U_{10}} - (w_2 + w_1) \times \text{Rate}_{U_{10}}],
\]

with arbitrary \( w_1 \) and \( w_2 \). These ICs are linear combinations of the disease odds, and under the rare-disease and no-redundancy assumptions, they are equal to linear combinations of the completion rates of the interaction classes \((U_7 \sim U_{12})\).

The coefficients in ICs can alternatively be expressed as the products of contrast coefficients for the gene \((G)\) and the environment \((E)\), respectively:

\[
\text{IC} = \sum_{i=1}^{3} \sum_{j=1}^{2} c_{iG} \times c_{jE} \times \text{Odds}_{s,1-i,j-1},
\]

where the genetic contrast coefficients \((c_{1G}, c_{2G}, c_{3G})\) are, respectively, \((-1, 1, 0)\) for \(\text{IC}_1\); \((-1, 0, 1)\) for \(\text{IC}_2\); and \((-w_1 - w_2, w_1, w_2)\) for \(\text{IC}_3\). The environmental contrast coefficients \((c_{1E}, c_{2E})\) are \((-1, 1)\) for all 3 ICs. As long as the sum-to-zero constraints,

\[
\sum_{i=1}^{3} c_{iG} = \sum_{j=1}^{2} c_{jE} = 0,
\]

are respected, an IC thus constructed will be equal to a linear combination of the completion rates of—and only of—the interaction classes. Clearly, a nonzero IC is mathematically incompatible with all interaction classes having a zero completion rate. A 2-sided test on an IC,

\[
\begin{cases}
H_0: \text{IC} = 0 \\
H_1: \text{IC} \neq 0,
\end{cases}
\]

is therefore a test for sufficient-cause gene-environment interaction.

The same principle applies in case-control studies, since the case-control odds estimated in a case-control study are a constant multiple (the reciprocal of the control sampling fraction of the study) of the corresponding disease odds in the underlying population (3). A straightforward way to estimate the case-control odds is to divide the number of cases (\(CS_{g,e}\)) by the corresponding number of controls (\(CN_{g,e}\)) in a stratum defined by genotype \((G = g)\) and environmental exposure level \((E = e)\) in a case-control data set, that is, \(\text{Odds}_{g,e} = \frac{CS_{g,e}}{CN_{g,e}}\). Web Appendix 1 (available at http://aje.oxfordjournals.org/) details the method for dealing with a polychotomous environmental exposure with a total of \(l\) levels and shows that as long as the sum-to-zero constraints for the contrast coefficients are respected, all of the resulting ICs are legitimate tests for sufficient-cause gene-environment interactions. Asymptotically, the test is a \(\chi^2\) test with 1 degree of freedom (df) for each constructed IC. One can also perform a single \(\chi^2\) test for several ICs simultaneously. The degrees of freedom equal the number of (linearly independent) ICs. For a ternary \(G\) and an \(l\)-leveled \(E\), the total number of degrees of freedom amenable for testing is \(2 \times (l - 1)\). A simultaneous test with the maximum number of degrees of freedom is referred to as the global test for sufficient-cause gene-environment interactions.

By judiciously designing the contrast coefficients, we can tailor an IC to detect a particular type of departure from additivity (sufficient-cause interactions) with greater sensitivity. For example, we can use \((c_{1G}, c_{2G}, c_{3G}) = (-1, 1/2, 1/2)\) for an autosomal dominant interaction, \((-1/2, -1/2, 1)\) for an autosomal recessive interaction, and \((-1, 0, 1)\) for a gene-dosage interaction. For \(l \geq 3\), there are also many possible choices for the environmental contrast coefficients, such as trend interaction (e.g., larger gene-environment interaction with a higher level of environmental exposure) or threshold interaction (e.g., gene-environment interaction occurs only when the environmental exposure reaches a certain threshold level). Specifying contrast coefficients as above, under the specific models they are designed to detect, will cause the ICs to deviate further from zero on average. However, a larger mean deviation is not enough; to be statistically relevant, we need the resulting IC to have a smaller variance as well. Web Appendix 2 shows a weighted version of the IC test, where the user-specified contrast coefficients are weighted by the inverse variances of the case-control odds.

### Imposing the independence and HWE assumptions

Because one of the 2 factors considered in this paper is genetic \((G)\) and the other is environmental \((E)\), we will next explore how to exploit the assumptions of GEI and HWE. If GEI can be assumed for the nondiseased subjects in the study population, a more efficient estimation for the case-control odds is

\[
\text{Odds}_{g,e}^{\text{GEI}} = \frac{CS_{g,e}}{\hat{u}_g \times CN_{g,e}^{+}},
\]

where \(\hat{u}_g = CN_{g,+}/CN_{+,+}\) is the estimate of the \(G = g\) genotype frequency in the nondiseased subjects in the study.
population and the “+” in the subscript indicates summation of the corresponding index.

If GEI and HWE are both assumed for the nondiseased subjects in the study population, the estimates for the case-control odds are

\[
\text{Odds}_{0,e}^{\text{GEI} \& \text{HWE}} = \frac{CS_{0,e}}{(1 - \hat{u})^2 \times CN_{+,e}},
\]

\[
\text{Odds}_{1,e}^{\text{GEI} \& \text{HWE}} = \frac{CS_{1,e}}{2 \times \hat{u} \times (1 - \hat{u}) \times CN_{+,e}},
\]

and

\[
\text{Odds}_{2,e}^{\text{GEI} \& \text{HWE}} = \frac{CS_{2,e}}{\hat{u} \times CN_{+,e}},
\]

respectively, where \(\hat{u} = (CN_{2,+} + 0.5 \times CN_{1,+})/CN_{+,+}\) is the estimated allele frequency among nondiseased subjects in the study population.

Web Appendix 3 presents the formulae of the asymptotic variances for all estimates. Web Appendix 4 presents the SAS code (SAS Institute, Inc., Cary, North Carolina) needed for all of the calculations. Users can specify genetic and environmental contrast coefficients to suit the most likely model of sufficient-cause interactions for the disease in question. Calculation of the weighted contrast coefficients is fully automatic, requiring no further input from the user. Web Appendix 5 presents an annotated example of such output.

**A SIMULATION STUDY**

We conducted a small-scale simulation study to examine the statistical properties of the proposed tests. We simulated data for a ternary gene (allele frequency 0.4) and a binary environmental exposure (prevalence 0.3), assuming GEI and HWE. For the null hypothesis of no sufficient-cause gene-environment interaction (\(H_0\)), we set the completion rates for all of the interaction classes (\(U_7 = U_{12}\)) to zero (Table 1). We further constructed a number of alternative hypotheses (Table 1). For the genetic factor, these include autosomal dominant (AD), autosomal recessive (AR), and gene-dosage (GD) interactions, respectively. For the environmental factor, all alternative hypotheses assumed an all-or-none (AN) interaction pattern (interaction for the exposed subjects but not for the unexposed subjects, completion rates for \(U_{10} \sim U_{12}\) being zero). For all hypotheses, the disease rates are on the order of 1 per 10,000 per year.

A case-control study with 1,000 cases and 1,000 controls was conducted in the study population. The proposed IC tests (both unweighted and weighted versions) were applied to the simulated data with various contrast coefficients, with or without the GEI and HWE assumptions. The \(\alpha\) level was set at 0.05. A total of 1,000,000 simulations were performed for each scenario. At this high number of simulations, the error is no more than \(\sqrt{(0.5 \times 0.5)/1,000,000} = 0.0005\).

Table 2 presents the type I error rates (under \(H_0\) in Table 1) of the IC tests and the weighted IC tests using different (specified) genetic contrast coefficients. The corresponding weighted genetic contrast coefficients are shown in the same rows as the specified ones. As noted above, the weighted coefficients are automatically calculated from the data once the unweighted ones are specified/supplied. Here we show the averages from the total 1,000,000 simulations. Type I error rates of the unweighted and weighted IC tests are well controlled for all of the contrast coefficients we specified, with or without imposing the assumptions of GEI and HWE. If one or both assumptions fail, only the unweighted tests and the weighted IC tests without the failed assumption(s) can maintain the nominal \(\alpha\) level (see Web Tables 1 and 2).

Table 3 presents the empirical powers of the IC tests and the weighted IC tests under the alternative hypothesis of an interaction between an autosomal dominant gene and an all-or-none environmental exposure (\(H_{\text{AD} \times \text{AN}}\) in Table 1). For all of the genetic contrast coefficients specified, statistical power increases when more assumptions (GEI and/or HWE) are imposed.

Table 3 also shows that given the same assumption(s), the power is highest for the 1-df weighted IC test with correctly specified genetic contrast coefficients; that is, the test using \((-0.78, 0.59, 0.19)\) that corresponds to the autosomal dominant coefficients \((-1, 1/2, 1/2)\). IC tests using contrast coefficients closer to \((-0.78, 0.59, 0.19)\) also have favorable powers—that is, 1) the unweighted IC test using \((-1, 1/2, 1/2)\) the

<table>
<thead>
<tr>
<th>Hypothesis</th>
<th>All-Unknown Class ((U_i))</th>
<th>Gene-Only Classes</th>
<th>Environment-Only Classes</th>
<th>Gene-Environment Interaction ((G \times E)) Classes</th>
</tr>
</thead>
<tbody>
<tr>
<td>(H_0)(^a)</td>
<td>1 4 2 1 3 1</td>
<td>0 0 0 0 0 0</td>
<td>0 0 0 0 0 0</td>
<td>0 0 0 0 0 0</td>
</tr>
<tr>
<td>(H_{\text{AD} \times \text{AN}})(^b)</td>
<td>1 4 2 1 3 1</td>
<td>4 4 1 0 0 0</td>
<td>0 0 0 0 0 0</td>
<td>0 0 0 0 0 0</td>
</tr>
<tr>
<td>(H_{\text{AR} \times \text{AN}})(^c)</td>
<td>1 4 2 1 3 1</td>
<td>6 1 1 0 0 0</td>
<td>0 0 0 0 0 0</td>
<td>0 0 0 0 0 0</td>
</tr>
<tr>
<td>(H_{\text{GD} \times \text{AN}})(^d)</td>
<td>1 4 2 1 3 1</td>
<td>6 3 1 0 0 0</td>
<td>0 0 0 0 0 0</td>
<td>0 0 0 0 0 0</td>
</tr>
</tbody>
</table>

**Abbreviations:** AD, autosomal dominant; AN, all or none; AR, autosomal recessive; GD, gene-dosage.

\(^a\) Null hypothesis.

\(^b\) AD for gene, AN for environmental exposure.

\(^c\) AR for gene, AN for environmental exposure.

\(^d\) GD for gene, AN for environmental exposure.
for HGD×AN). The global test can detect all types of alternatives.

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examined the relationship between polymorphisms in the Table 3.

interaction subspace, regardless of the values used for the weights. This is because the pair of vectors spans the entire interaction subspace, regardless of the values used for the weights.

Table 2. Genetic Contrast Coefficients and Type I Error Rates (Under H\textsubscript{0} in Table 1) of the Interaction Contrast Tests, for the Data Simulated Under the Assumptions of Gene-Environment Independence and Hardy-Weinberg Equilibrium

<table>
<thead>
<tr>
<th>Type of Interaction More Sensitive to Detection</th>
<th>Specified</th>
<th>Weighted</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>G = 0</td>
<td>G = 1</td>
</tr>
<tr>
<td></td>
<td>G = 0</td>
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</tr>
<tr>
<td>Autosomal dominant</td>
<td>–1</td>
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</tr>
<tr>
<td>Autosomal recessive</td>
<td>–1/2</td>
<td>–1/2</td>
</tr>
<tr>
<td>Gene-dosage</td>
<td>–1</td>
<td>0</td>
</tr>
<tr>
<td>Global</td>
<td>–1</td>
<td>1</td>
</tr>
<tr>
<td>–1</td>
<td>0</td>
<td>1</td>
</tr>
</tbody>
</table>

The table shows the genetic contrast coefficients and type I error rates for different types of genetic interactions under the assumptions of gene-environment independence and Hardy-Weinberg equilibrium.

<table>
<thead>
<tr>
<th>Type of Interaction</th>
<th>Genetic Contrast Coefficienta</th>
<th>Type I Error Rate</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Assuming Neither GEI nor HWE</td>
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</tr>
<tr>
<td></td>
<td>IC Test</td>
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</tr>
<tr>
<td>–1</td>
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<td>1</td>
</tr>
</tbody>
</table>

Abbreviations: GEI, gene-environment independence; HWE, Hardy-Weinberg equilibrium; IC, interaction contrast.

a The environmental contrast coefficients are specified as (–1, 1) for (E = 0, E = 1) for all scenarios, with the corresponding weighted environmental contrast coefficients calculated as (–0.71, 0.71).

weighted IC test using (–0.81, 0.33, 0.48) that corresponds to the gene-dosage contrast coefficients (–1, 0, 1). The weighted and unweighted tests are the same for the 2-df global interaction test. This is because the pair of vectors spans the entire interaction subspace, regardless of the values used for the weights.

The 1-df weighted IC test with correctly specified genetic contrast coefficients has the highest power to detect the other 2 alternatives as well (Web Table 3 for H\textsubscript{AR×AN}, Web Table 4 for H\textsubscript{GDN×AN}). The global test can detect all types of alternatives listed in Table 1 with reasonably high power (Table 3, Web Tables 3 and 4). We also tested other allele frequencies and exposure prevalences and achieved similar results (not shown). Based on the simulation results, we recommend using the 1-df weighted IC test when one has a priori knowledge about the interaction model and using the global test if one does not.

AN EXAMPLE

We used Sam et al.’s (21) case-control data on upper aerodigestive tract cancers as an example. In this study, Sam et al. examined the relationship between polymorphisms in the cytochrome P-450, family 1, subfamily A, polypeptide 1 gene (CYP1A1)—CYP1A1*2A and CYP1A1*1A, with the former being the variant allele—and smoking (yes/no) in the risk of contracting upper aerodigestive tract cancers (Table 4). None of the cells in Table 4 have counts of less than 5. The assumptions of GEI (P = 0.7563) and HWE (P = 0.6789) are both tenable among control subjects. Lee et al. (9) previously analyzed these data assuming a multiplicative model for disease odds. Under that model, they concluded that the (multiplicative) interactions between CYP1A1 polymorphisms and smoking were nonsignificant at an α level of 0.05, with or without the assumptions of GEI and HWE.

We next applied the weighted IC test to these data for any sufficient-cause gene-environment interaction (Table 5). Without the assumptions of GEI and HWE, none of the genetic contrast coefficients we tried (autosomal dominant, autosomal recessive, gene-dosage, and global) resulted in statistically significant test results. However, with the GEI assumption invoked, the weighted IC tests became significant at α = 0.05 for all contrasts. When both the GEI assumption and the HWE assumption were invoked, all 3 sets of genetic

Table 3. Genetic Contrast Coefficients and Empirical Powers (Under H\textsubscript{AD×AN} in Table 1) of the Interaction Contrast Tests, for the Data Simulated Under the Assumptions of Gene-Environment Independence and Hardy-Weinberg Equilibrium

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provide at most 6 gene- and environmental exposure–specific disease rates (the data degrees of freedom). An estimation of model parameters is not possible in a nonidentifiable model, but hypothesis testing is a different story. This paper demonstrates that the IC test can detect gene-environment interactions, even under the nonidentifiable sufficient-cause model. The significance of an IC test implies the presence of at least 1 interaction class involved in the given contrast; that is, some sufficient-cause gene-environment interaction is occurring. A nonsignificant test result, however, does not guarantee the opposite; a perfect cancellation of several interaction classes with nonzero completion rates also leads to IC = 0. An example is the “leaky” (L) environmental exposure where the interaction completion rates for the unexposed subjects (U_{10}, U_{11}, U_{12}) are exactly 1 \times 10^{-5} shy of those for the exposed subjects (U_{7}, U_{8}, U_{9}) in Table 1. Here, we have no power whatsoever against H_{AD}L, H_{AR}L, and H_{GD}L alternatives. This is an unavoidable limitation for any hypothesis testing of the parameters of a nonidentifiable model.

The rare-disease assumption is necessary for the proposed method, under which a sufficient-cause model for rates is equivalent to a sufficient-cause model for odds. Web Table 5 presents the biases of using odds to approximate rates. The biases are less than 0.05% for rates under 0.001 per year—the setting for studies of cancers, coronary heart diseases, etc. For more common diseases, such as hypertension or type 2 diabetes, the approximation breaks down, and the method proposed here would be inapplicable.

Another necessary assumption, the no-redundancy assumption, is more subtle and is not amenable to testing by itself. This is actually a much weaker assumption than the simple independent action assumption, which originated from toxicopharmacology (23) and found use in epidemiology in recent decades (17, 20, 24–31). In the language of the sufficient-cause model, the simple independent action assumption posits that the arrival events of the unknown complement causes in different classes of sufficient causes are independent of one another. The no-redundancy assumption can still hold, even if there is strong dependency in the arrival events (and the simple independent action assumption fails).

### Table 4. A Case-Control Data Set From the Study by Sam et al. (21) and the Estimated Case-Control Odds

<table>
<thead>
<tr>
<th>No. of Cases</th>
<th>No. of Controls</th>
<th>Estimated Case-Control Odds^a</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Assuming Neither GEI nor HWE</td>
</tr>
<tr>
<td>---------------</td>
<td>----------------</td>
<td>--------------------------------</td>
</tr>
<tr>
<td>E = 0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>G = 0</td>
<td>55</td>
<td>0.79</td>
</tr>
<tr>
<td>G = 1</td>
<td>76</td>
<td>1.23</td>
</tr>
<tr>
<td>G = 2</td>
<td>19</td>
<td>2.11</td>
</tr>
<tr>
<td>E = 1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>G = 0</td>
<td>91</td>
<td>2.02</td>
</tr>
<tr>
<td>G = 1</td>
<td>123</td>
<td>4.24</td>
</tr>
<tr>
<td>G = 2</td>
<td>44</td>
<td>8.80</td>
</tr>
</tbody>
</table>

Abbreviations: GEI, gene-environment independence; HWE, Hardy-Weinberg equilibrium.

^a The probability of a study subject’s being a case divided by the probability of his/her being a control in a case-control study.

### Table 5. Genetic Contrast Coefficients and P Values for the Data Shown in Table 4

<table>
<thead>
<tr>
<th>Type of Interaction</th>
<th>Genetic Contrast Coefficient^a</th>
<th>P Value From the Weighted Interaction Contrast Test</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Specified</td>
<td>Weighted</td>
</tr>
<tr>
<td></td>
<td>G = 0</td>
<td>G = 1</td>
</tr>
<tr>
<td>Autosomal dominant</td>
<td>–1</td>
<td>1/2</td>
</tr>
<tr>
<td>Autosomal recessive</td>
<td>–1/2</td>
<td>–1/2</td>
</tr>
<tr>
<td>Gene-dosage</td>
<td>–1</td>
<td>0</td>
</tr>
<tr>
<td>Global</td>
<td>–1</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>–1</td>
<td>0</td>
</tr>
</tbody>
</table>

Abbreviations: GEI, gene-environment independence; HWE, Hardy-Weinberg equilibrium.

^a The environmental contrast coefficients are specified as (−1, 1) for (E = 0, E = 1) for all scenarios, with the corresponding weighted environmental contrast coefficients calculated as (−0.71, 0.71).

^b Significant at a false discovery rate of 5%.

^c Significant at α = 0.05 after Bonferroni correction.
To break the no-redundancy assumption, the unknown complement causes for 2 different classes need to have a common constituent factor, and that common factor must be the last one to arrive among all of the constituents of these 2 classes, before all other classes are completed.

The remaining 2 assumptions, the GEI and HWE assumptions, are optional, and when their validity is in doubt, a researcher always has the liberty to put them to test using the data at hand. Han et al. (32) exploited the independence assumption to develop a constrained likelihood ratio test for gene-environment interactions under an additive risk model. However, their method cannot take into account the HWE assumption, and it is highly demanding computationally. By contrast, all the formulae presented here—with or without the GEI and HWE assumptions—are of the closed form, obviating the need for a computer-intensive iteration algorithm. Lee et al.’s (9) modified logistic regression model can exploit both assumptions and can be easily fitted using common statistical packages. However, it detects multiplicative interactions, not sufficient-cause gene-environment interactions, which are the focus of this paper.

While the proposed IC test can be tailored to detect a particular type of sufficient-cause gene-environment interaction with greater sensitivity, it is actually a nonspecific test. To pin down a specific interaction—for example, the $U_7$ interaction class—one can test for max(Odds$S_{2,1} - Odds_{0,1} - Odds_{1,0}$, Odds$_{2,1} - Odds_{1,1} - Odds_{2,0} > 0$ (33–35)). VanderWeele referred to this as a “singular interaction” or an “epistatic interaction,” since there are individuals for whom the outcome would occur if and only if $G = g$ and $E = e$ (33–35). This is because Odds$_{2,1} - Odds_{0,1} - Odds_{2,0} = Rate_{U_7} - Rate_{U_1} - Rate_{U_4} - Rate_{U_{14}}$ and Odds$_{2,1} - Odds_{1,1} - Odds_{2,0} = Rate_{U_7} - Rate_{U_1} - Rate_{U_2} - Rate_{U_{12}}$, so if statistically either one is larger than zero, it must be the case that Rate$_{U_i} > 0$, and therefore the presence of the $U_7$ interaction class can be inferred. Further work is needed to develop empirical tests for these special forms of gene-environment interaction in case-control studies, with and without the assumptions of GEI and HWE.

Besides the genetic and environmental factors under consideration, there may be other factors that could confound gene-environment interactions. It may also be that the study population is not a homogeneous one but instead is composed of several population strata (36–38). The assumptions of GEI and HWE hold within each population stratum but do not hold in the population as a whole. To account for these, one can stratify the data according to confounders and population strata and then perform a separate IC test in each resulting stratum. With a proper multiple-testing correction for multiple strata, the presence of some sufficient-cause gene-environment interaction can be inferred if the result of any of these stratum-specific IC tests turns out to be significant. Further work is needed to develop stratified sufficient-cause interaction testing methods, both when the total number of strata is large (and the average stratum size is small, i.e., the sparse-data scenario) and when some of the stratifying variables interact with the specific gene and environmental exposure under study (sufficient-cause interactions between 3 or more variables).

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REFERENCES


